

Multiplex TEMPase 2x Master Mix

3 mM MgCl₂ final concentration



Cat. No.: 733-2569 (2500 Reactions)

Cat. No.	No. of 50 µl Reactions	Multiplex TEMPase 2x Master Mix	MgCl ₂ 25 mM
ID No. Cap colour	-	5500100 Green	5575801 Yellow
733- 2570 Sample	20	1 x 0.5 ml	1 x 1.5 ml
733- 2417	100	2 x 1.25 ml	1 x 1.5 ml
733- 2568	500	10 x 1.25 ml	1 x 1.5 ml
733- 2569	2500	50 x 1.25 ml	3 x 1.5 ml

Store at -20 °C.

Key Features

- Amplification of multiple DNA targets in one tube
- High specificity, sensitivity and product yield
- Diminished formation of non-specific products
- Detection of low copy number targets

Multiplex PCR is a method that enables amplification of two or more amplicons simultaneously in a single reaction tube. It is widely used in genotyping and different areas of DNA testing in research, forensic and diagnostic laboratories.

Multiplex TEMPase 2x Master Mix facilitates the amplification of multiple PCR products, minimizes the need for optimization, making the development of multiplex PCR assays both fast and simple.

Multiplex TEMPase 2x Master Mix is an all-in-one 2x master mix with TEMPase Hot Start DNA polymerase, optimized buffer system, dNTPs and MgCl₂. Each reaction requires 25 µl of the 2x Master Mix. Simply add primers, template and water to a total reaction volume of 50 µl to successfully carry out multiplex PCR.

TEMPase Hot Start DNA Polymerase is a modified form of VWR Taq DNA Polymerase, which is activated by heat treatment. A chemical moiety is attached to the enzyme at the active site, which renders the enzyme inactive at room temperature. Thus, during setup and the first ramp of thermal cycling, the enzyme is not active and misprimed primers are not extended. The result is higher specificity and greater yields when compared to standard DNA polymerases.

Kit Components

Multiplex TEMPase 2x Master Mix

TEMPase Hot Start DNA polymerase, optimized buffer system, dNTPs and 6 mM mMgCl₂

MgCl₂

25 mM MgCl₂ in PCR grade water

Recommended Storage and Stability

Long term storage at -20 °C. Product expiry at -20 °C is stated on the label.

Option: Store at +4 °C for up to 6 months.

Quality Control

TEMPase Hot Start DNA Polymerase is tested for contaminating activities, with no traces of endonuclease activity, nicking activity or exonuclease activity.

Unit Definition

One unit is defined as the amount of polymerase that incorporates 10 nmoles of dNTPs into acid-precipitable DNA in 30 minutes at 72°C under standard assay conditions.

Protocol

This protocol serves as a guideline for PCR. Optimal reaction conditions such as incubation times, temperatures, and amount of template DNA may vary and must be determined individually.

Notes:

- All primers should have the same melting temperature.
- Use equal concentrations (0.2 µM) of all primers for the initial test.
- If one product gives a much stronger band than the others, reduce the primer concentration for this target.
- If one product gives a much weaker band than the others, increase the primer concentration for this target.
- Set up reaction mixtures in an area separate from that used for DNA preparation or product analysis. Working on ice is not required.
- The MgCl₂ concentration in the final reaction is 3 mM with this Master Mix. In some applications, more MgCl₂ is required for best results. Use 25 mM MgCl₂ to adjust the MgCl₂ concentration according to table 1.

Table 1. Additional volume (µl) of MgCl₂ per 50 µl reaction:

Final MgCl ₂ conc. in reaction (mM)	3.0	3.5	4.0	4.5	5.0
Volume of 25 mM MgCl ₂	0	1	2	3	4

1. Thaw Master Mix and primers. **It is important to thaw the solutions completely and mix thoroughly before use to avoid localized concentrations of salts.**

Important: Spin vials briefly before use.

2. Prepare the reaction mix. Table 2 shows the reaction mix set-up for one reaction with a final volume of 50 µl.
3. Mix the reaction mix thoroughly and dispense appropriate volumes into reaction tubes.

Table 2. Reaction mix and template DNA

Component	Vol./reaction*	Final concentration*
Master Mix	25 µl	1x
25 mM MgCl ₂	0 µl (0 – 7 µl)	3 mM (0.5 – 5 mM)
Each forward Primer (10 µM)	1 µl (0.5 – 5 µl)	0.2 µM (0.1 – 1.0 µM)
Each reverse primer (10 µM)	1 µl (0.5 – 5 µl)	0.2 µM (0.1 – 1.0 µM)
PCR-grade H ₂ O	X µl	-
Template DNA	X µl	genomic DNA: 200 ng (10 – 500 ng) plasmid DNA: 0.5 ng (0.1 – 1 ng) bacterial DNA: 5 ng (1 – 10 ng)
TOTAL volume	50 µl	-

* Suggested starting conditions; theoretically used conditions in brackets

- Add template DNA to the individual tubes containing the reaction mix.
- Program the thermal cycler according to the manufacturer's instructions. **Each program must start with an initial heat activation step at 95°C for 15 minutes.** See table 3 for an example.
For maximum yield and specificity, temperatures and cycling times should be optimized for each new template or primer pair.
- Place the tubes in the thermal cycler and start the reaction.

Table 3. Three-step PCR program

Cycles	Duration of cycle	Temperature
1	15 minutes ^a	95 °C
30 – 40	20 – 30 seconds ^b 50 – 90 seconds ^c 50 – 90 seconds ^d	95 °C 50 – 65 °C 72 °C
1	4 minutes ^e	72 °C

^a. For activation of the TEMPase hot start enzyme.

^b. Denaturation step: This step is the first regular cycling event and consists of heating the reaction to 95 °C for 20 – 30 seconds. It causes melting of the DNA template by disrupting the hydrogen bonds between complementary bases, yielding single-stranded DNA molecules.

^c. Annealing step: The reaction temperature is lowered to 50 – 65 °C for 20 – 40 seconds allowing annealing of the primers to the single-stranded DNA template. Typically, the annealing temperature is about 3 – 5 °C below the T_m (melting temperature) of the primers used.

^d. Extension/elongation step: TEMPase polymerase has its optimum activity temperature at 72 °C. At this step the DNA polymerase synthesizes a new DNA strand complementary to the DNA template strand. The extension time depends on the length of the DNA fragment to be amplified. As a rule of thumb, at its optimum temperature the DNA polymerase will polymerize a thousand bases per minute.

^e. Final elongation: This single step is occasionally performed at a temperature of 72 °C for 5 minutes after the last PCR cycle to ensure that any remaining single-stranded DNA is fully extended.

Related Products

Description	Cat. No.
VWR Taq DNA Polymerase	
VWR Glycerol Free Taq DNA Polymerase (1000 units) with 10x Key Buffer with 10x Extra Buffer	733-1817
VWR Taq DNA Polymerase (500 units) with 10x Key Buffer with 10x Extra Buffer	733-1301
VWR Taq DNA Polymerase (500 units) with 10x Extra Buffer (Mg ²⁺ -free and Triton-free)	733-1304
VWR Taq DNA Polymerase (1000 units) with 10x Key Buffer (Tween-free) with 10x Extra Buffer (Triton-free)	733-1307
VWR Taq DNA Polymerase (500 units) with 10x Key Buffer (Mg ²⁺ -free)	733-1311
VWR Red Taq DNA Polymerase (2500 units) with 10x Key Buffer with 10x Extra Buffer	733-1323
VWR Taq DNA Polymerase Master Mix*	
VWR Taq DNA Pol. 1.1x Master Mix, 1.5 mM MgCl ₂ final conc. (2500 reactions)	733-1314
VWR Taq DNA Pol. 2x Master Mix, 1.5 mM MgCl ₂ final conc. (2500 reactions)	733-1316
VWR Red Taq DNA Pol. 1.1x Master Mix, 1.5 mM MgCl ₂ final conc. (2500 reactions)	733-1318
VWR Red Taq DNA Pol. 2x Master Mix, 1.5 mM MgCl ₂ final conc. (2500 reactions)	733-1320
VWR TEMPase Hot Start DNA Polymerase	
VWR TEMPase Hot Start DNA Polymerase (500 units) with 10x Key Buffer with 10x Combination Buffer	733-1331
VWR Glycerol Free TEMPase DNA Polymerase (500 units) with 10x Key Buffer	733-2555
VWR TEMPase Hot Start Master Mix**	
VWR TEMPase Hot Start 2x Master Mix K, 1.5 mM MgCl ₂ final conc. (2500 reactions)	733-2582
VWR Blue TEMPase Hot Start 2x Master Mix K, 1.5 mM MgCl ₂ final conc. (2500 reactions)	733-2585
VWR Multiplex Master Mix	
VWR Multiplex 2x Master Mix, (2500 reactions)	733-2569
VWR High Fidelity DNA Polymerase	
VWR AccuPOL DNA Polymerase (250 units) with 10x Key Buffer	733-1324
VWR AccuPOL DNA Polymerase (250 units) with 10x Key Buffer (Mg ²⁺ free and Tween free)	733-1328
VWR PCR accessories	
Betaine Enhancer Solution 5 M, 5 ml (500 reactions)	733-1361
dNTP Mix, 40 mM (10 mM of each dATP, dCTP, dGTP & dTTP) (40 µmol, 2x 500 µl)	733-1363
dNTP Set, 100 mM of each dATP, dCTP, dGTP & dTTP (4x 25 µmol, 4x 250 µl)	733-1364
VWR PCR grade water (6 x 5 ml)	733-2573
VWR Loading Buffer Red (5 x 1 ml) – available in 4 different colors	733-2574

*Also available with 2 mM MgCl₂

**Also available as 2x Master Mix C based on Combination Buffer

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