

CB-XTM **Protein Assay**

One Assay for All Jobs

INTRODUCTION

Protein assays are routinely used in many research fields to estimate proteins in a vast array of buffers and conditions. A major problem for researchers is to select a protein assay from the vast selection on the market that is compatible with their protein sample. $CB-X^{TM}$ Protein Assay eliminates this problem as it is designed to be compatible with all commonly used buffers and conditions in protein isolation, storage and assays.

For protein samples in simple, uncomplicated aqueous buffers $CB-X^{TM}$ is a highly sensitive, single reagent assay that can be performed in 5 minutes. $CB-X^{TM}$ Protein Assay uses a protein dye that is an improvement on the Bradford Coomassie dye reagent.

For complicated protein samples CB-X[™] Protein Assay is supplied with reagents to clean up the samples and remove all reagents and chemicals that interfere with accurate protein estimation. These reagents include detergents, chaotropes, reducing agents, alkylating agents, sugars, high salt concentrations, buffering agents and chelating agents (Table 1). The clean up stage and subsequent protein assay is performed in a single tube to ensure no protein loss and to maintain the accuracy of the assay.

Figure 1 depicts the simple scheme of the CB-XTM Protein Assay. If the protein sample does not contain interfering agents then a straightforward single reagent assay is performed to give a linear response. If interfering agents are present or if an artifact results are produced then the protein samples are treated with the clean up reagents and the protein is then assayed generating a linear response.

CB-XTM Protein Assay is supplied with a lot specific CB-XTM Tables. These allow researchers to perform single protein clean ups, subsequent assays and then look up their absorbance in the CB-XTM Table to find the protein concentration. The CB-XTM Table eliminates the need for multiple protein standards and saves considerable time and effort. The CB-XTM Table is prepared with a complex protein mixture that compares well with proteins from mammalian, plant, bacteria and yeast sources. A set of bovine serum albumin standards are supplied for generating curves when using CB-XTM Assay Dye alone or for researcher's who prefer to generate their own standard curve or to generate their own CB-XTM Table for their specific conditions.



Figure 1: CB-X[™] Protein Assay Scheme. Protein samples are rapidly assayed with a single reagent with in five minutes. In most cases the assay will generate accurate results, however if interfering agents are present artifactual results may occur. In this situation the samples are cleaned with a one-step procedure in a single tube. This generates accurate protein estimations.

DETERGENTS		REDUCING AGENTS	
Brij [®] 35	2%	2-mercaptoethanol	1M
CHAPS	2%	DTT	1M
CHAPSO	2%	CHAOTROPES	
Nonidet [®] P-40	2%	Guanidine.HCl	6M
SDS	2%	Urea	6M
Triton [®] X-100	2%	SALTS	
Tween [®] 20	2%	Ammonium sulfate	1M
Deoxycholate	0.1%	MISCELLANEOUS	
SUGARS		EDTA	0.1M
Glucose	1M	HEPES	0.1M
Sucrose	25%	MES	0.1M

 Table 1: CB-X[™] Protein Assay is compatible with many interfering agents

The CB-XTM Protein Assay is reliable over the range of $0.5-50\mu g$ per assay. The regular size kit contains enough CB-XTM Assay Dye for 500 protein assays and enough clean up reagents for 250 clean ups. (*Patents Pending*)



ITEM(S) SUPPLIED

Cat. #	786-12X	786-12XT
CB-X TM	2 x 125ml	10ml
CB-X [™] Assay Dye	2 x 250ml	10ml
CB-X [™] Solubilization Buffer-I	15ml	1ml
CB-X [™] Solubilization Buffer-II	15ml	1ml
CB-X Protein Standard (2mg/ml)	5ml	-
$CB-X^{TM}$ Table: Lot Specific	1	1

STORAGE CONDITION:

The kit is shipped at ambient temperature. Upon arrival, store CB-X[™] Assay Dye and bovine serum albumin (BSA) standard at 4°C and other kit components at room temperature. The CB-X[™] reagent must be chilled to -20°C for optimal efficiency, for additional convenience the CB-X[™] reagent can be stored at -20°C. When stored and used properly this kit is good for 12 months.

ITEMS NEEDED AND NOT SUPPLIED WITH THIS KIT:

- Centrifuge
- Assay tubes (G-Biosciences Cat # 786-008)
- Disposable polystyrene cuvettes (*G-Biosciences Cat* # 786-009)

PREPARATIONS BEFORE USE:

- Chill CB-X[™] at -20°C. *NOTE: CB-X[™] can be stored at -20°C for prolonged storage and additional convenience.* Prior to use, mix CB-X[™] Assay Dye by gently inverting the bottle several times. DO NOT SHAKE THE BOTTLE.

PROTOCOL 1: FOR SIMPLE, UNCOMPLICATED AQUEOUS SAMPLES

- 1. Dilute the supplied BSA Standard in the same buffer as your protein samples. We recommend duplicates of 5-6 dilutions within the range of 0.1-1.0mg/ml.
- 2. Transfer 50μ l diluted protein standards and test samples to the assay tubes. Set up a blank containing 50μ l of the buffer used to generate the standards.
- 3. Add 1ml CB-X[™] Assay Dye into each tube and mix well. Incubate for 5 minutes at room temperature.
- 4. Read the absorbance at 595nm.
- 5. Prepare a standard calibration plot for the determination of protein concentration of the unknown samples.

NOTE: This assay can be performed in titer plates. Use 10µl protein standards or samples and 200µl $CB-X^{TM}$ Assay Dye and follow the above protocol.

PROTOCOL 2: FOR INTERFERING AGENT REMOVAL (SAMPLE CLEAN UP) & PROTEIN ESTIMATION

- 1. Transfer 5-100µl protein sample to a 1.5ml centrifuge tube.
- 2. Add 1ml *pre-chilled* (-20°*C*) CB-XTM and vortex to mix.
- 3. Centrifuge at 16,000xg for 5 minutes and carefully remove all the supernatant without disturbing the protein pellet.
- 4. Add 50µl CB-X[™] Solubilization Buffer-I and 50µl CB-X[™] Solubilization Buffer-II to the tube and vortex to dissolve the protein pellet. NOTE: Most proteins will quickly dissolve; however insoluble and membrane proteins may take 2-10 minutes of periodic vortexing.
- 5. Invert the CB-X[™] Assay Dye 2-3 times to mix and add 1ml CB-X[™] Assay Dye to the tube and vortex briefly. Incubate for 5 minutes at room temperature.

- 6. Read the absorbance at 595nm against deionized water using either a 1cm path length cuvette or transfer 200µl assay solution to a microtiter well.
- 7. Use the appropriate CB-X[™] Table to determine the amount of protein in your sample. For cuvettes use the CB-X[™] Table for Spectrophotometer and for microtiter wells use the CB-X[™] Table for Microplate Reader.
- 8. Calculate the protein concentration $(\mu g/\mu l)$ by dividing of the amount of protein (μg) by the sample volume (μl) .

PROTOCOL OPTIMIZATION

For routine protein assays, we recommend that researchers generate their own $CB-X^{\mathbb{M}}$ Tables or standard curves. This is a one time commitment to ensure user specific results and allows single tube assays to be performed each time as opposed to generating new calibration plots for each assay. We recommend using a protein standard similar to you protein of interest or a purified source of your protein.

The supplied $CB-X^{\text{TM}}$ tables may result in some inconsistencies due to the type of cuvettes or microtiter plates used. $CB-X^{\text{TM}}$ Table for Spectrophotometer readings were measured with 1 cm path length cuvettes against deionized water and $CB-X^{\text{TM}}$ Table for Microplate Reader readings were measured using NuncTM Immuno 96 MicroWellTM Plates (MaxiSorpTM) [Cat# 442404] against deionized water.

- 1. Prepare duplicate standards of choice in your buffer of choice at the following concentrations: 0, 0.2, 0.4, 0.6, 0.8, $1.0 \mu g/\mu l$.
- 2. Transfer 50µl protein standard to 1.5ml centrifuge tubes and following the Protocol 2 from step 2 to 6.
- 3. Prepare a standard calibration plot for the determination of protein concentration of the unknown samples. Use the line equation to generate your own CB-X[™] Tables.
- 4. This CB-X[™] Table will allow all your future protein estimations to be performed without using protein standards, allowing you to carry out rapid, single protein estimations. NOTE: If your assay conditions change a new custom CB-X[™] Table will need to be generated.

RELATED PRODUCTS

1. <u>Non-Interfering Protein AssayTM (Cat # 786-005)</u>

*NI-Protein Assay*TM is not affected by interfering agents commonly present in protein solutions, including reducing agents such as 2*ME*, *DTT*, detergents, amines, *EDTA*, salts, sugars, etc. *NI-Protein Assay*TM also shows no protein-to-protein variation. The assay is based on removal of interfering agents prior to assay by a single step protocol. Sensitivity as low as 0.5ug/assay. Assay time is 15-20 minutes

2. <u>dotMETRIC[™] - 1 µl Protein Assay (Cat # 786-20/21)</u>

For sample economy and rapid estimation of protein using a test strip. Simply apply 1μ protein solution on the test strip, develop test strips in 8 minutes, and measure the diameter of protein spot on test strip for determination of protein concentration. dotMETRICTM assay is resistant to reducing agents, detergents and shows little or no protein to protein variation.

3. Bovine Gamma Globulin Standard (2mg/ml) (Cat # 786-007) – Protein standard for protein estimation.

NOTE: For other related products, visit our web site at <u>www.GBiosciences.com</u> or contact us.