ICEme™
MX-ICP Mutation Enrichment Kit

Limitations of ICEme Kit

Contaminating substances, carried over from extraction of formalin-fixed paraffin-embedded samples, may interfere with the MX PCR amplification of the samples. The quality control procedures outlined in the ICEme Kit User Guide will ensure that the extracted DNA is suitable for use in this kit.

This kit has been tested on DNA isolated from cell lines, formalin-fixed paraffin-embedded tumor samples and plasma. It is a **RESEARCH USE ONLY KIT** and is not for diagnostic use.

Care must be taken to avoid carryover and cross-contamination with this kit. The extreme sensitivity of the assay method requires precautions to be taken at the following points:

- Ensure that all samples are handled such that cross-contamination between samples cannot occur.
  - It is recommended to wipe down each vial with 10% bleach prior to opening, however care must be taken to ensure that the bleach solution does not get into the solution contained in the vials.
- Work in a PCR workstation or other suitable space where the work area can be wiped down with a 10% bleach solution and exposed to UV light (20 – 30 min) prior to setting up PCR amplification reactions.
  - After using the PCR workstation or hood it is recommended to wipe down the surfaces with a 10% bleach solution and expose the area to UV light.

If ANY solutions such as molecular biology grade water are to be kept in the PCR workstation or hood, they must be kept in glass containers as repeated exposure to UV light will break down plastic and the breakdown products will be present in the solution and inhibit the PCR reactions.

Except for pipette tips, any other plastic container used in making the master mixes or in setting up the reactions (96-well plates, caps, or strip tubes) should NOT be kept in the hood and exposed to repeated cycles of UV light as the plastic breaks down and the breakdown products will inhibit the PCR reactions.

- Use a separate PCR workstation or room for opening the samples after MX PCR amplification for Quality Control by gel electrophoresis.
- Ensure that the kit's Positive Control is handled separately from test samples at all stages of the assay.
  - Make sure all solutions, no–template-controls and sample DNA wells are capped prior to opening the tubes containing the Positive Control.
  - HANDLE THE POSITIVE CONTROL LAST. Add the Positive Control to the appropriate tubes only AFTER ALL No-Template-Control and Test Sample wells have been capped.
  - After capping the Positive Control tube, wipe ALL tubes and caps with a DNA destroying agent (such as 10% bleach) prior to transfer to another area.
  - Note: when a 10% bleach solution is used, it must be made up fresh each week in a dark/opaque bottle.
  - Note: 10% (v/v) bleach is used for destroying DNA under 500 bases in length.
  - Note: High molecular weight DNA must be destroyed using a commercially available agent for DNA decontamination.
- Ensure that when pipetting samples into 96-well plates, you do not allow sample contamination of adjacent wells either due to splashing during mixing or by not changing pipette tips.

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