



# $XIT^{^{\text{TM}}}$ Genomic DNA Blood Kits

For the isolation of genomic DNA from fresh blood, bone marrow and buffy coat

# INTRODUCTION

The XIT™ Genomic DNA Blood kits are designed for the isolation of genomic DNA from whole blood, bone marrow and buffy coat. The XIT<sup>™</sup> kit uses the principle of cell lysis, protein precipitation and finally DNA precipitation to isolate high quality genomic DNA.

XIT Genomic DNA Blood Kit protocol is designed to use 0.5ml whole blood, however the protocol can be easily adapted for larger tissue sample sizes. The purified DNA has an A260/A280 ratio between 1.8 - 2.0 and has yields ranging between 10-15µg/ml depending on volume of blood.

	Cat # 786-294	Cat # 786-295	Cat # 786-296
ITEM(S) SUPPLIED	Up to 12.5ml blood	Up to 125ml blood	Up to 250ml blood
RBC Lysis Buffer	100ml	2 x 250ml	4 x 250ml
XIT <sup>™</sup> Lysis Buffer	10ml	100ml	2 x 100ml
XIT <sup>™</sup> Protein Precipitation Buffer	2.5ml	25ml	2 x 25ml
TE Buffer	1.5ml	20ml	2 x 20ml
LongLife <sup>™</sup> RNase	0.5ml	0.5ml	2 x 0.5ml

## STORAGE CONDITIONS

The kit is shipped at ambient temperature. Upon arrival, store the  $LongLife^{TM}$  Proteinase K and  $LongLife^{TM}$  RNase at -20°C and all other kit components at room temperature. The kit components are stable for 1 year, if stored properly.

# ITEMS NEEDED BUT NOT SUPPLIED

- Isopropanol
- 70% ethanol

#### PREPARATION BEFORE USE

1. Preheat a water-bath or heating block to 55°C and equilibrate TE Buffer to 50-60°C.

# **PROTOCOL**

- 1. For processing buffy coats, use the volumes required for processing the original blood sample. For example, if the buffy coat preparation was processed from 5ml whole blood then follow the Protocol for 5ml Blood.
- For bone marrow samples, ensure that the sample is completely homogenous after addition of XIT<sup>™</sup> Lysis Buffer. If not add additional XIT<sup>™</sup> Lysis Buffer until an homogenous sample is obtained.

## FOR 0.5ml BLOOD

- 1. Add 0.5ml whole blood to a 1.5ml tube containing 1ml RBC Lysis Buffer. Invert the tube to mix and incubate 2-3 minutes at room temperature.
- Centrifuge 14,000xg for 30 seconds then remove supernatant carefully without disturbing the pellet.
- Add 1ml of RBC Lysis Buffer to the pellet and mix.
- Centrifuge 14,000xg for 30 seconds then remove supernatant. Repeat step 3 and 4 if pellet is not white.
- Vortex the tube to resuspend the cells in the residual liquid.



6. Add 400µl of XIT<sup>™</sup> Lysis Buffer to the resuspended cells and vortex vigorously to lyse the cells. Usually no incubation is required; however, if cell clumps are visible after mixing, incubate at 37°C for 5-10 minutes or until the solution is homogenous.

<u>OPTIONAL</u>: Add  $2\mu l$  LongLife<sup>TM</sup> RNase solution to the cell lysate, mix by inverting the tube 10-15 times and incubate at  $37^{\circ}C$  for 15 minutes.

- 7. Add 90µ1 XIT<sup>™</sup> Protein Precipitation Buffer to the sample and mix by inverting the tube 10-20 times.
- 8. Centrifuge at 16,000g for 5 minutes. Carefully, transfer the supernatant to a new tube.

NOTE: The supernatant should be clear. If not, repeat the centrifugation.

- 9. Add 400µl isopropanol to the supernatant and mix by gently inverting the sample at least 20-25 times.
- 10. Centrifuge at 14,000rpm for 5 minutes.
- 11. Discard the supernatant and use a pipette to carefully remove remaining liquid without disturbing the DNA pellet.
- 12. Add  $200\mu l$  70% ethanol and invert the tube twice to wash the pellet.
- 13. Centrifuge at 14,000rpm for 5 minutes.
- 14. Discard the supernatant and drain the tube on a piece of clean absorbent paper. Allow to air dry for 15 minutes.
- 15. Add 50µl TE buffer to dissolve the DNA.
- 16. Rehydrate the genomic DNA by incubating at 55-65°C for one hour, followed by an overnight incubation at room temperature to ensure complete genomic DNA hydration.
- 17. Store DNA at 4°C, for long term storage store at -20 or -80°C.

### PROTOCOL FOR 5ml BLOOD

- 1. Add 5ml whole blood to a 15ml centrifuge tube containing 5ml RBC Lysis Buffer. Invert the tube to mix and incubate 2-3 minutes at room temperature.
- 2. Centrifuge 2,000xg for 5 minutes then remove supernatant carefully without disturbing the pellet.
- 3. Add 10ml of RBC Lysis Buffer to the pellet and mix.
- 4. Centrifuge 2,000xg for 5 minutes then remove supernatant. Repeat step 3 and 4 if pellet is not white.
- 5. Vortex the tube to resuspend the cells in the residual liquid.
- 6. Add 4ml of *XIT*<sup>™</sup> Lysis Buffer to the resuspended cells and vortex vigorously to lyse the cells. Usually no incubation is required; however, if cell clumps are visible after mixing, incubate at 37°C for 5-10 minutes or until the solution is homogenous.

<u>OPTIONAL</u>: Add  $20\mu l$  LongLife<sup>TM</sup> RNase solution to the cell lysate, mix by inverting the tube 10-15 times and incubate at  $37^{\circ}C$  for 15 minutes.

- 7. Add 900µl XIT<sup>TM</sup> Protein Precipitation Buffer to the sample and mix by inverting the tube 10-20 times.
- 8. Centrifuge at 2,000g for 5 minutes. Carefully, transfer the supernatant to a new tube.

**NOTE**: The supernatant should be clear. If not, repeat the centrifugation.

- 9. Add 4ml isopropanol to the supernatant and mix by gently inverting the sample at least 20-25 times.
- 10. Centrifuge at 2,000g for 5 minutes.
- 11. Discard the supernatant and use a pipette to carefully remove remaining liquid without disturbing the DNA pellet.
- 12. Add 2ml 70% ethanol and invert the tube twice to wash the pellet.
- 13. Centrifuge at 2,000g for 3 minutes.
- 14. Discard the supernatant and drain the tube on a piece of clean absorbent paper. Allow to air dry for 15 minutes.
- 15. Add 500µl TE buffer to dissolve the DNA.

- 16. Rehydrate the genomic DNA by incubating at 55-65°C for one hour, followed by an overnight incubation at room temperature to ensure complete genomic DNA hydration.
- 17. Store DNA at 4°C, for long term storage store at -20 or -80°C.

# **RELATED PRODUCTS**

- EZ-Grind<sup>™</sup> (Cat # 786-139): A highly efficient grinding resin that is pre-aliquot into 1.5ml grinding tubes and is supplied with matching pestles.
- 2. <u>Pestle & Tubes (Cat. #786-138P):</u> DNase/RNase free microfuge tubes (1.5ml) and matching pestles for the grinding of small samples and isolation of nuclei.
- 3. <u>Molecular Grinding Resin™ (Cat # 786-138):</u> For grinding of small samples. High tensile micro particles that do not bind nucleic acids, allowing most samples to be processed by hand using inexpensive micro centrifuge tube pestles or a mortar and pestle.

**NOTE:** For other related products, visit our web site at <u>www.GBiosciences.com</u> or contact us.