

FOR FAECAL CONCENTRATION OF

HELMINTH OVA AND LARVAE / PROTOZOA CYSTS AND OOCYSTS



Midi Parasep[®] SI

Health and Safety Benefits

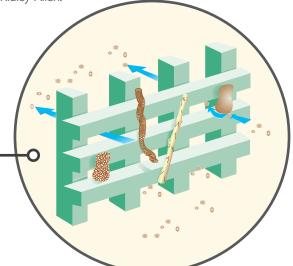
- Totally enclosed/sealed process
- Solvent free protocol
- Disposable device
- Single use, no sample contamination
- Ready to use systems available

Performance Benefits

- Optimum sample recovery
- Enhanced sample clarity
- Rapid four step process
- Human resources optimised
- Reduces human error

Patented Filter (High Density Polyethylene)

A two stage filtration matrix. Large particles are rejected without occluding the 425µm pores. Recovery rate with Parasep[®] is comparable to traditional sieve method, ie: Ridley-Allen.



Debris Trap

Rejected particles are trapped to prevent extrusion into the Sedimentation Cone during centrifugation.

Air/Liquid Seal and Safety Lock

The 'seal' prevents the release of biohazardous material. The 'lock' ensures the Mixing Chamber and Filter are removed together for safe disposal.

Fat Dispersion Chamber

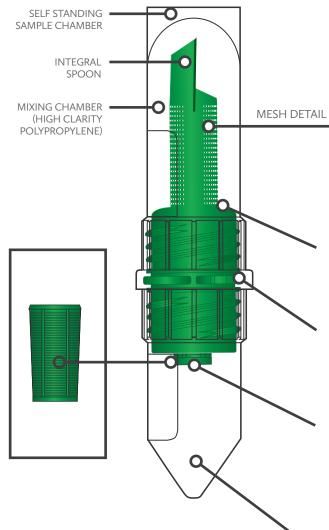
A perforated fat dispersion chamber (EC Des.App.000512017) removes the smaller faecal debris and separates the fat content so that it can be efficiently removed from the resulting sediment without the use of ether or ethyl acetate.

Sedimentation Cone

Parasites are efficiently collected in the sedimentation cone during centrifugation and easily aspirated/ pipetted for microscopic examination.

Centrifuge Compatibility

Designed to fit all 50ml centrifuge buckets.



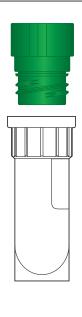
Procedure

See label for storage conditions and expiry date. Please adhere to the following guidelines when handling Midi Parasep[®] SF. To avoid cross contamination the Midi Parasep[®] SF device should remain closed at all times except when introducing the sample or when retrieving the final concentrated sample for examination.

STEP 1 - SAMPLE PREPARATION

Preserved Samples

Transfer 3ml of the emulsified surfactant treated stool into the mixing chamber.



STEP 2 - EMULSIFICATION

Seal Midi Parasep® SF by screwing in the filter/sedimentation cone unit. Vortex or shake to emulsify with the sedimentation cone pointing upwards.



STEP 3 - CENTRIFUGATION

Invert the Midi Parasep® SF and centrifuge at 400g for two minutes.

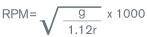
Midi Parasep® SF fits all 50ml centrifuge buckets.

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DEBRIS .

SEDIMENT -

NOTE: TO CALCULATE THE REQUIRED RPM FOR ANY CENTRIFUGE.



RPM - rotor speed in revs/min. g - centrifugal force (max.1000g) r - radius, horizontal distance between sedimentation cone tip and spindle centre measured in mm.

STEP 4 - EXAMINATION

Direct Method

Unscrew and discard the filter and mixing tube. Pour off all the liquid above the sediment.

From the resulting sediment you can conduct both wet mounts and permanent staining methods.

For wet mounts, pipette one drop of Saline or lodine solution onto a slide, add one drop of deposit to the Saline or lodine, mix sample and cover with cover-slip.

OR

ParaSys™

Unscrew and discard the filter and mixing tube. Pour off all the liquid above the sediment.

Press 'Dilute' to add Saline to the sediment. Shake or vortex to re-suspend sample. Insert Aspirator into suspension and press SAMPLE to draw 100µl into the ParaSlide[™] . (Refer to ParaSys[™] instruction manual). SOLVENT FREE FAECAL PARASITE CONCENTRATOF



Ordering Information

MIDI F	PARASEP [®] SF	STORAGE TEMP	PACK SIZE	VWR Cat No.
Midi Par	asep® SF (Devices supplied without reagent)	4-35°C	50	10048-714

Please refer to the product labelling and pack insert for details of the relevant hazard information.

